

**TRANSMITTAL LETTER TO THE UNITED STATES
DESIGNATED/ELECTED OFFICE (DO/EO/US)
CONCERNING A SUBMISSION UNDER 35 U.S.C. 371**

ATTORNEY DOCKET NUMBER
59866.000004

U.S. APPLICATION NO. (if known, see 37 CFR 1.31)

Unassigned

INTERNATIONAL APPLICATION NO.
PCT/DK2003/000634

INTERNATIONAL FILING DATE
26 September 2003

PRIORITY DATE CLAIMED
26 September 2002

TITLE OF INVENTION

A METHOD FOR DETECTING, SCREENING AND/OR MONITORING A CANCER IN AN INDIVIDUAL

APPLICANT(S) FOR DO/EO/US

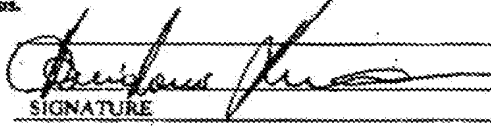
Lasse L. HESSEL, Jesper MALLING, Nils BRÜNNER, Mads HOLTEN-ANDERSEN, Hans Jørgen NIELSEN

Applicant herewith submits to the United States Designated/Elected Office (DO/EO/US) the following items and other information:

1. ☒ This is a **FIRST** submission of items concerning a filing under 35 U.S.C. 371.
2. ☐ This is a **SECOND** or **SUBSEQUENT** submission of items concerning a filing under 35 U.S.C. 371.
3. ☐ This expresses request to begin national examination procedures (35 U.S.C. 371(f)). The submission must include items (5), (6), (9) and (21) indicated below.
4. ☒ The US has been elected (Article 31).
5. ☒ A copy of the International Application as filed (35 U.S.C. 371(c)(2))
 - a. ☒ is attached hereto (required only if not communicated by the International Bureau).
 - b. ☒ has been communicated by the International Bureau.
 - c. ☐ is not required, as the application was filed in the United States Receiving Office (RO/US).
6. ☐ An English language translation of the International Application as filed (35 U.S.C. 371(c)(2)).
 - a. ☐ is attached hereto.
 - b. ☐ has been previously submitted under 35 U.S.C. 154(d)(4).
7. ☐ Amendments to the claims of the International Application under PCT Article 19 (35 U.S.C. 371(c)(3))
 - a. ☐ are attached hereto (required only if not communicated by the International Bureau).
 - b. ☐ have been communicated by the International Bureau.
 - c. ☐ have not been made; however, the time limit for making such amendments has **NOT** expired.
 - d. ☐ have not been made and will not be made.
8. ☐ An English language translation of the amendments to the claims under PCT Article 19 (35 U.S.C. 371(c)(3)).
9. ☒ An unexecuted oath or declaration of the inventor(s) (35 U.S.C. 371(c)(4)).
10. ☐ An English language translation of the annexes to the International Preliminary Examination Report under PCT Article 36 (35 U.S.C. 371(c)(5)).

Items 11 to 20 below concern document(s) or information included:

11. ☐ An Information Disclosure Statement under 37 CFR 1.97 and 1.98.
12. ☐ An Assignment document for recording. A separate cover sheet in compliance with 37 CFR 3.28 and 3.31 is included.
13. ☒ A Preliminary Amendment.
14. ☒ An Application Data Sheet under 37 CFR 1.76.
15. ☐ A substitute specification.
16. ☐ A power of attorney and/or change of address letter.
17. ☐ A computer-readable form of the sequence listing in accordance with PCT Rule 13ter.2 and 37 CFR 1.821-1.825.
18. ☐ A second copy of the published International Application under 35 U.S.C. 154(d)(4).
19. ☐ A second copy of the English language translation of the international application under 35 U.S.C. 154(d)(4).
20. ☐ Other items or information:

U.S. APPLICATION NO. IF KNOWN, SEE 37 CFR 1.51		INTERNATIONAL APPLICATION NO. PCT/DK2003/000634		ATTORNEY'S DOCKET NUMBER 59866.000004	
21. <input checked="" type="checkbox"/> The following fees are submitted:				CALCULATIONS PTO USE ONLY	
<input checked="" type="checkbox"/> a) Basic National Fee		\$ 300.00	\$	300.00	
<input checked="" type="checkbox"/> b) Examination Fee		\$ 200.00	\$	200.00	
<input checked="" type="checkbox"/> c) Search Fee		\$ 500.00	\$	500.00	
TOTAL OF ABOVE CALCULATIONS		\$1000.00	\$	1000.00	
<input type="checkbox"/> Additional fee for specification and drawings filed in paper over 100 sheets (excluding sequence listing or computer program listing filed in an electronic medium). The fee is \$250.00 for each additional 50 sheets of paper or fraction.					
Total Sheets	Extra Sheets	Number of each additional 50 or fraction thereof (round up to a whole number)	RATE		
- 100	/50		x \$250.00	\$	
Surcharge of \$130.00 for furnishing the oath or declaration later than 30 months from the earliest claimed priority date (37 CFR 1.492(e)).				\$	
CLAIMS	NUMBER FILED	NUMBER EXTRA	RATE	\$	
Total claims	26 - 20 =	6	x \$ 50.00	\$	300.00
Independent claims	1 - 3 =	0	x \$200.00	\$	
MULTIPLE DEPENDENT CLAIM(S) (if applicable)			+ \$360.00	\$	
TOTAL OF ABOVE CALCULATIONS =				\$	1,300.00
<input checked="" type="checkbox"/> Applicant claims small entity status. See 37 CFR 1.27. The fees indicated above are reduced by 1/2.				\$	650.00
SUBTOTAL =				\$	650.00
Processing fee of \$130.00 for furnishing the English translation later than 30 months from the earliest claimed priority date (37 CFR 1.492(f)).				\$	
TOTAL NATIONAL FEE =				\$	650.00
Fee for recording the enclosed assignment (37 CFR 1.21(h)). The assignment must be accompanied by an appropriate cover sheet (37 CFR 3.28, 3.31). \$40.00 per property.				\$	
TOTAL FEES ENCLOSED =				\$	650.00
				Amount to be refunded:	\$
				Amount to be charged:	\$
a. <input type="checkbox"/> A check in the amount of _____ to cover the above fees is enclosed. b. <input checked="" type="checkbox"/> Please charge Deposit Account No. <u>50-0206</u> in the amount of <u>\$625.00</u> to cover the above fees. A duplicate copy of this sheet is enclosed. c. <input checked="" type="checkbox"/> The Commissioner is hereby authorized to charge any additional fees which may be required, or credit any overpayment to Deposit Account No. <u>50-0206</u> . A duplicate copy of this sheet is enclosed. d. <input type="checkbox"/> Fees are to be charged to a credit card. WARNING: Information on this form may become public. Credit card information should not be included on this form. Provide credit card information and authorization on PTO-2038.					
NOTE: Where an appropriate time limit under 37 CFR 1.495 has not been met, a petition to revive (37 CFR 1.137(a) or (b)) must be filed and granted to restore the application to pending status. SEND ALL CORRESPONDENCE TO:					
CUSTOMER NO. 21967 Intellectual Property Department Huron & Williams LLP 1900 K Street, N.W., Suite 1200 Washington, DC 20006-1109 (202) 955-1500 (telephone) (202) 778-2201 (facsimile)			 SIGNATURE Stanislaus Aksman NAME 28,562 REGISTRATION NUMBER		
			March 28, 2003 DATE		



PATENT
ATTY. DOCKET NO. 59866.000004

IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

Application Number : TBA
Applicant : Lasse L. HESSEL et al.
Filed : March 28, 2005
Title : A METHOD FOR DETECTING, SCREENING AND/OR
MONITORING A CANCER IN AN INDIVIDUAL
TC/Art Unit : TBA
Examiner: TBA
Docket No. : 59866.000004
Customer No. : 21967

Confirmation No.: TBA

PRELIMINARY AMENDMENT UNDER 37 C.F.R. § 1.115

MAIL STOP AMENDMENT
Commissioner for Patents
P.O. Box 1450
Alexandria, VA 22313-1450

Sir:

Prior to examination on the merits, please amend the above-identified patent application as follows:

Amendments to the Specification begin on page 2 of this paper.

Amendments to the Claims are reflected in the listing of claims which begins on page 4 of this paper.

Remarks begin on page 8 of this paper.

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AMENDMENTS TO THE SPECIFICATION:

Please add the new paragraph on page 1, line 3, after the title, "A METHOD FOR DETECTING, SCREENING AND/OR MONITORING A CANCER IN AN INDIVIDUAL".

CROSS-REFERENCE TO RELATED APPLICATIONS

This patent application is a U.S. national phase application of PCT/DK/2003/000634, filed September 26, 2003, which claims priority to Denmark Patent Application No. 2002 01430, filed September 26, 2002, both of which are incorporated herein by reference in their entireties. This patent application is also a continuation-in-part of U.S. patent application 09/546,573, filed April 10, 2000 ("573 application"), and a continuation-in-part of U.S. patent application 10/117,030, filed April 8, 2002 ("030 application"). The '573 and '030 applications claim priority to Denmark Patent Application No. DK 1999 00476 filed April 9, 1999 ("DK '0476 application"). The entire contents of the '573, '030 and DK '0476 applications are incorporated herein by reference in their entireties.

Please insert the following new paragraph at page 8, before line 4, as indicated below:

One embodiment of the invention is directed to a method for detecting a cancer in an individual, screening an individual for cancer, monitoring an individual for cancer, or the combination of the detecting, the screening and the monitoring an individual for cancer. The method comprises determining a first parameter represented by a concentration of TIMP-1 in at least one excreta from the individual. The presence of the first parameter above a predetermined discrimination value is an indication that the individual has a likelihood (such as a high likelihood) of having the cancer or that there is a progression of the cancer in the individual.

Please delete the paragraph 3, at page 9, lines 14-17, and insert the replacement paragraph as indicated below:

Such cancers could comprise, e.g., be breast, prostate, colorectal, cervical, ovarian, lung, pancreatic, renal, vulvar, hepatocellular carcinomas, minimal residual disease and recurrent cancer. In this application whenever a group

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of elements is recited with the transitional phrase "comprising", it is understood that we also contemplate the same group of elements with transitional phrases "consisting essentially of", "consisting", or with the phrase "selected from the group of consisting of" preceding the recitation of the elements. Thus, such cancers are, e.g., selected from the group consisting of breast carcinoma, prostate carcinoma, colorectal carcinoma, cervical carcinoma, ovarian carcinoma, lung carcinoma, pancreatic carcinoma, renal carcinoma, vulvar carcinoma, hepatocellular carcinoma, minimal residual disease and recurrent cancer, and, optionally, mixtures thereof.

AMENDMENTS TO THE CLAIMS:

Please amend claims 3-12, 15-20, and 22 as follows.

1. (Original) A method for detecting and/or screening and/or monitoring a cancer in an individual, said method comprising determining a first parameter represented by the concentration of TIMP-1 in at least one excreta from the individual, wherein the presence of the first parameter above a predetermined discrimination value is an indication that the individual has a high likelihood of either having a cancer or progression in a cancer.
2. (Currently Amended) A method according to claim 1 wherein the cancer is selected from the group comprising consisting of breast carcinoma, prostate carcinoma, colorectal carcinoma, cervical carcinoma, ovarian carcinoma, lung carcinoma, pancreatic carcinoma, renal carcinoma, vulvar carcinoma, and hepatocellular carcinomas, minimal residual disease and recurrent cancer.
3. (Currently Amended) A method according to ~~any of the preceding~~ claims, claim 1, wherein the excreta is saliva.
4. (Currently Amended) A method according to claim 1, ~~2 or 3~~, wherein the first parameter is the total concentration of TIMP-1.
5. (Currently Amended) A method according to claim 1, ~~2 or 3~~, wherein the first parameter is the combination of the concentration of total TIMP-1 and the concentration of free TIMP-1.
6. (Original) A method according to claim 5, wherein the combination is performed by logistic regression analysis.
7. (Currently Amended) A method according to ~~any of the preceding~~ claims, claim 1, wherein the discrimination value is determined by determining the total concentration of TIMP-1 in the at least one excreta in both a healthy control

population and a population with known cancer, thereby determining the discriminating discrimination value which identifies the cancer population with a predetermined specificity or a predetermined sensitivity.

8. (Currently Amended) A method according to any of the preceding claims, claim 1, wherein the method further comprises determining at least one second parameter representing the concentration of a marker for cancer different from any form of TIMP-1, in an excreta from an individual.

9. (Currently Amended) A method according to claim 8, wherein the first and second parameter are combined to result in a combined parameter wherein the presence of a concentration of the combined parameter above a predetermined discrimination value is an indication that the individual has a high likelihood of having a cancer or that there is ~~being~~ a progression in a cancer.

10. (Currently Amended) A method according to claim 9, wherein the predetermined discrimination value is determined by determining the combined parameter in the at least one excreta in both a healthy control population and a population with known colorectal cancer, thereby determining the predetermined discriminating discrimination value which identifies the cancer population with a predetermined specificity or a predetermined sensitivity.

11. (Currently Amended) A method according to claim 9 or 10, wherein the combination of the first and second parameter is performed by logistic regression analysis.

12. (Currently Amended) A method according to ~~claim 8-11~~ claim 8, wherein the at least one second parameter is the concentration of Carcino Embryonic Antigen (CEA).

13. (Currently Amended) A method according to ~~any of the preceding claims,~~ claim 7, wherein the determination of the total concentration of TIMP-1 is performed by means of an immunoassay or an active assay.
14. (Currently Amended) A method according to ~~any of the preceding claims,~~ claim 13, wherein the immunoassay is an ELISA.
15. (Currently Amended) A method according to ~~any of the preceding claims,~~ claim 13, wherein the active assay is zymography.
16. (Currently Amended) Use of a A method according to claim 1-45, for ~~which detection detects~~ of early stage cancer.
17. (Currently Amended) Use A method according to claim 16, for ~~which detection detects~~ of early stage colorectal cancer.
18. (Currently Amended) Use A method according to claim 16, for ~~which detection detects~~ of metastatic breast cancer.
19. (Currently Amended) Use of a A method according to claim 1-45, for ~~which monitoring monitors~~ the response to cancer treatment.
20. (Currently Amended) Use of a A method according to claim 1-45, for ~~which monitoring monitors~~ the recurrence of a cancer.
21. (Currently Amended) A dipstick for performing the method according to claim 1-45, wherein said dipstick comprises a first colour indication zone, comprising antibodies specific for TIMP-1.
22. (Currently Amended) A dipstick according to claim 21, wherein the first zone further comprises at least one reagent which ~~can gives~~ an optically visible

colour change in the zone dependent on the concentration of TIMP-1 in at least one excreta.

23. (Currently Amended) A dipstick according to claim 21 or 22, wherein the dipstick further comprises a second colour indication zone, able to react with at least one substance normally present in the excreta, and thereby providing an optically visible colour change in the zone for controlling that proper use of the dipstick stick is used properly.

24. (Currently Amended) A dipstick according to claim 24-23, wherein said dipstick further comprises a third colour indication zone, comprising antibodies specific for CEA.

25. (Currently Amended) A dipstick according to claim 24-24, wherein the first zone further comprises at least one reagent which can give an optically visible colour change in the zone dependent on the concentration of CEA in at least one excreta.

26. (New) A method according to claim 1 wherein the cancer comprises breast carcinoma, prostate carcinoma, cervical carcinoma, ovarian carcinoma, lung carcinoma, pancreatic carcinoma, renal carcinoma, vulvar carcinoma, hepatocellular carcinoma, minimal residual disease or recurrent cancer.

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REMARKS

The specification has been amended to update a cross-reference to related patent applications reflecting priority claims. Applicants respectfully submit that this amendment to the cross-reference to related applications is submitted within the time period set forth in 37 C.F.R. § 1.78 for claiming benefit of earlier filing dates.

The specification has also been amended to add a description of various aspects of Applicants' invention present in other parts of the application as filed. Thus, the amendments find written description and enablement in the application, considered as a whole, e.g., in the original claims 1-2.

Claim amendments are supported by the specification, including original claims, considered as a whole.

An indication of allowance of all claims is respectfully requested.

Authorization is hereby granted to charge or credit the undersigned's Deposit Account No. 50-0206 for any fees or overpayments related to the entry of this Amendment.

APPLN. NO. TBA
PRELIMINARY AMDT. DATED MARCH 28, 2005
ATTORNEY DOCKET NO.: 59866.000004

Respectfully submitted,

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By: 

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Dated: March 28, 2005

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APPLICATION DATA SHEET

Application Information

Application Number::	To Be Assigned
Filing Date::	3/28/05
Application Type::	Regular
Subject Matter::	Utility
Suggested Classification::	
Suggested Group Art Unit::	
CD-ROM or CD-R::	None
Number of CD Disks::	
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Sequence Submission?::	
Computer Readable Form (CRF)?::	
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Title Line One::	A METHOD FOR DETECTING, SCREENING AND/OR MONITORING A CANCER IN AN INDIVIDUAL
Title Line Two::	
Attorney Docket Number::	59866.000004
Request for Early Publication?::	No
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Suggested Drawing Figure::	
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Small Entity?::	Yes
Petition Included?::	No
Petition Type::	
Licensed US Government Agency::	
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Representative Information

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Domestic Priority Information

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This Application is	a National Phase of	PCT/DK2003/000634	September 26, 2003
This Application is	a CIP of	09/546,573	April 10, 2000
This Application is	a CIP of	10/117,030	April 8, 2002

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Country:	Application Number:	Filing Date:	Priority Claimed:
Denmark	200201430	September 26, 2002	Yes
Denmark	199900476	April 9, 1999	Yes

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HU, ID, IL, IN, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LR,
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Declarations under Rule 4.17:

- as to the identity of the inventor (Rule 4.17(i)) for the fol-
lowing designations AE, AG, AL, AM, AT, AU, AZ, BA, BB,
BG, BR, BY, BZ, CA, CH, CN, CO, CR, CU, CZ, DE, DK,
DM, DZ, EC, EE, EG, ES, FI, GB, GD, GE, GH, GM, HR,
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SD, SL, SZ, TZ, UG, ZM, ZW), Eurasian patent (AM, AZ,
BY, KG, KZ, MD, RU, TJ, TM), European patent (AT, BE,
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IE, IT, LU, MC, NL, PT, RO, SE, SI, SK, TR), OAPI patent
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SN, TD, TG)
- of inventorship (Rule 4.17(iv)) for US only

Published:

- with international search report
— before the expiration of the time limit for amending the
claims and to be republished in the event of receipt of
amendments

For two-letter codes and other abbreviations, refer to the "Guid-
ance Notes on Codes and Abbreviations" appearing at the begin-
ning of each regular issue of the PCT Gazette.

(54) Title: A METHOD FOR DETECTING, SCREENING AND/OR MONITORING A CANCER IN AN INDIVIDUAL

(57) Abstract: The invention relates to a method for screening and/or detecting and/or monitoring a cancer in an individual, said method comprising determining a first parameter represented by the concentration of TIMP-1 in at least one excreta, e.g. saliva, from the individual. The invention provides a method that without the need to use a blood sample is suitable for facilitating the early diagnosis of a cancer, monitoring the recurrence of a cancer, and/or monitoring the status of a cancer or the effect of cancer treatment in an individual.

WO 2004/029627 A1

The present invention relates to a method for early detection and/or screening and/or monitoring of a cancer in an individual.

15 Furthermore, if a cancer is detected, an early detection of the spread of cancer, the recurrence of cancer after treatment as well as the response to treatment, is crucial to the survival of cancer patients.

Colorectal cancer (CRC) is currently the fourth most common cancer in the US and ranks second as a cause of cancer-related deaths. Evidence exists that reductions in colorectal cancer morbidity and mortality can be achieved through detection and treatment of early-stage colorectal cancers and the identification and removal of adenomatous polyps, the precursors of colorectal cancer.

Currently, the only way of diagnosing colorectal cancer is to carry out a colonoscopy, which is impractical as a screening technique for general applicability. Furthermore, since the disease is still primarily managed by surgical resection, there has been great interest in screening.

A known screening test for colorectal cancer that is often recommended on a routine basis by many health-care organizations is the fecal occult blood test (FOBT), which uses various methods to detect blood in the stool. However, although the test has a high specificity, this method has an intermediate sensitivity often leading to either false-positive or false-negative results, and may therefore cause unwarranted anxiety about cancer and unnecessary further tests or may miss the disease in its early stages.

The American Society of Clinical Oncology (ASCO) has therefore recommended against the use of fecal occult blood tests as a means for regular postoperative monitoring of colorectal cancer, (Benson et al, JCO, 2000). ASCO has also emphasised that new surveillance methods for the detection of colorectal cancer recurrences are needed (Benson et al, JCO, 2000).

Because metastatic disease is the main cause of cancer patient morbidity and mortality, molecules involved in the regulation of tumour invasion and metastasis are attractive as potential screening/detection/monitoring targets. It is well established that proteolytic enzymes produced by cancer cells or by cells in the tumour stroma are involved in extracellular tissue degradation, leading to cancer cell invasion and metastasis. A number of enzymes have been associated with this process, the most thoroughly investigated being the metalloproteinases, such as the collagenases and stromelysins, and the serine proteases such as plasmin. Recently, data has been published indicating that these molecules, free or bound in complexes, are released from tumour tissue and find their way into the circulation.

A positive correlation between tumour cell aggressiveness and matrix metalloproteinases (MMPs) expression has been well documented. MMPs are a group of zinc-binding endopeptidases involved in connective tissue matrix remodelling and degradation of the extracellular matrix (ECM), which are essential steps in tumour invasion, angiogenesis, and metastasis (reviewed by Aznavoorian et al, 1993).

However, matrix metalloproteinase enzyme activity is highly regulated. Regulation occurs at multiple levels, including modulation of MMP gene expression, extracellular activation of the pro-enzyme, inhibition of activation and enzymes activity by association with tissue inhibitors of metalloproteinase (TIMPs) (Matrisian, 1990), (Mauviel, 1993), (Birkedal-Hansen, 1993).

TIMPs forms stoichiometric complexes with both latent and active MMP's (Welgus et al., 1985; Kleiner et al., 1993), thereby inhibiting the catalytic activity of these enzymes (Stetler-Stevenson et al., 1996; Goldberg et al., 1992; Birkedal-Hansen et al., 1993).

TIMPs represent a family of ubiquitous proteins. There have, at present, been identified four members of the TIMP family. It has been found that TIMP-1 and TIMP-2 are capable of inhibiting tumour growth, invasion, and metastasis (Liotta, et al., 1991 and Khokha et al., 1989). However, recent clinical studies have stated TIMP-1 and TIMP-2 to be rather tumour-promoting molecules, as they were found to be significantly overexpressed in patients with poor prognosis. (McCarthy et al, 1999; Remacle et al, 2000).

Studies have shown that TIMP-1 in plasma is a highly sensitive and specific screening marker for the identification of patients with a high risk of having colorectal cancer (Holten-Andersen et al., 1999 and Holten-Andersen et al. 2002).

A method using this information is disclosed in WO00/62070, the complete disclosure of which is incorporated herein by reference for all purposes. In said reference it has been found that individuals can be screened for colorectal cancer by measuring the TIMP-1 concentration in plasma samples.

However, the method has the drawback that a blood sample must be drawn for each individual. This can be extremely expensive and troublesome when detecting and monitoring cancers in large populations and in some cases even impossible e.g. in the developing countries.

Furthermore, trained staff must draw blood samples, and particular care must be taken with the samples, as there is a high prevalence of hepatitis B and the human immunodeficiency virus (HIV) infections in some populations, e.g. in the developing countries.

Based on these problems other fluids mentioned in the exhausted list of WO/0062070 such as urine and serum has been evaluated as an alternative to plasma, as these are the traditional biological samples for the qualitative and quantitative measurement of substances including proteins.

However, serum studies have shown that there is no significant difference between the total TIMP-1 concentrations in serum samples from individuals having colorectal cancer and healthy blood donors, most properly because the platelets will leak TIMP-1 during coagulation in vitro (Cooper et al, 1985; Jung et al, 1996). Furthermore, in other studies, TIMP-1 has by itself been proven inadequate as a serum marker for screening for pancreatic cancer, (Zhou et al, 1998).

The concentration of TIMP-1 in urine and plasma from the same colorectal cancer patients has also been evaluated, but without

finding any noticeable correlation, since no TIMP-1 elevation was found in the urine (Brünner, unpublished data).

It is known for the person skilled in the art, that all organic compounds of plasma, such as hormones, immunoglobulines, enzymes, proteins, DNA and viruses may be detected in saliva in trace amounts (Vining and McGinley, 1985). However, it is a well established knowledge that the concentration found in saliva is lower than the concentration in urine which again is lower than the concentration found in plasma.

The lower concentrations in saliva are caused by a number of factors. As an example can be mentioned that saliva contains enzymes for digestion and that the total protein concentration in saliva is negligible since it is less than 1% of that in plasma (Breimer and Danhof, 1980). Furthermore it is likely that a major source of these trace amounts originates from the gingival crevicular fluid (from the tooth/gum margin) (Cimasoni, 1974).

A person skilled in the art would therefore expect that the natural enzymes, such as proteases in the saliva, would digest these negligible protein concentrations and reduce any concentration found in e.g. saliva to a negligible concentration if detectable at all.

One of a number of examples of a protein having a lower concentration in urine than in serum/plasma and again an even lower concentration in saliva, is prostate-specific antigen (PSA), which is the most commonly used serum marker for screening for prostate cancer, (Balk et al, 2003). PSA is present in high concentrations in serum and plasma, in minor concentrations in urine, and in one study no detectable PSA levels have been found in saliva at all, (Lovgren et al, 1999). Furthermore, another study has shown that determination of free and total PSA in saliva to improve and simplify the

differentiation between prostate cancer and benign prostatic hyperplasia is not suitable for use as an alternative measurement of serum (Turan et al, 2000).

5 Accordingly, since no elevated levels of TIMP-1 could be found in urine of either healthy blood donors or individuals suffering from colorectal cancer, the person skilled in the art would based on the common general knowledge that the concentration of a protein in saliva always would be lower than
10 in urine and plasma have no hint to examine saliva as a possible body fluid for detection of TIMP-1.

It should in this respect be noted, that TIMP-1 can be found in sputum from patients with asthma and chronic bronchitis
15 (Vigonola et al, 1998), but TIMP-1 is in such cases coming from blood from small lesions in e.g. the lung tissue.

Thus, as the current available screening methods for colorectal cancer are based on blood or stool samples, there is a long
20 felt need for a method for detecting, screening and monitoring cancers, which is applicable for large populations without the need to use blood or stool samples.

It is therefore one aspect of the present invention to provide
25 a screening method, which is suitable to facilitate the early diagnosis of a cancer without the need to use blood or stool samples.

It is another aspect of the present invention to provide a
30 method for monitoring the recurrence of a cancer, status of a cancer or the effect of cancer treatment in an individual, without the need to use a blood or stool sample.

It is a third aspect of the present invention to provide a
35 device for performing the method according to the invention, having a simple and inexpensive design, being quick and easy to

use and not requiring the assistance of specialists or the use of specialised equipment.

This is achieved according to the invention as it has surprisingly been found by the inventors that the TIMP-1 protein can be detected in human excreta, such as saliva in unexpected high concentrations with respect to the TIMP-1 concentration in plasma.

10 The inventors hereby overcome the common prejudice that a protein cannot be detected in saliva if it has not been detected in urine. As previously stated, earlier studies have shown, that noticeable concentrations of TIMP-1 protein could not be detected in urine and there has therefore been no indication in the prior art for the skilled person to look for TIMP-1 in saliva.

Furthermore, as a non-invasive technique, the collection and analysis of the excreta would appear to be particularly attractive for a high-risk patient population where the routine collecting of blood is often made difficult because of bruised or thrombosed veins.

20 In addition, this would be beneficial to both the patient (blood sampling would be reduced) and to those who handle patient samples (clinic and laboratory staff) e.g. because saliva can inhibit HIV infectivity (Wolff and Hay, 1991).

30 It is therefore possible according to the present invention to provide an easy and inexpensive method for early detection and monitoring of a cancer in an individual or large populations using a non-invasive technique for collection of samples.

In order not to obtain any false-positive results it is important to determine a discriminating value, which divides

the tested individuals in a group having either a high or low likelihood of having cancer.

It has been found that in one embodiment of the invention the total concentration of TIMP-1 is measured, that is, the sum of the TIMP-1 in free form and the TIMP-1 in complex forms. The person skilled in the art will understand that other expressions than the exact concentration can represent the concentration, such as, e.g., the concentration multiplied by a factor, etc., and that such other representations can be used equally well for the purpose of the present invention provided the corresponding adjustments are made.

The discriminating value is established by measuring the total concentration of TIMP-1 in both a healthy control population and a population with known cancer and thereby determining the discriminating value. The discriminating value identifies the cancer population with either a predetermined specificity or a predetermined sensitivity or both, and is based on an analysis of the relation between the concentration values and the known clinical data of the healthy control population and the cancer patient population.

The discriminating value determined in this manner is valid for the same experimental set-up in future individual tests.

Specificity is defined as the proportion of non-diseased individuals having a parameter representing the concentration of a marker, such as TIMP-1 in excreta samples lower than a predefined level.

Sensitivity is defined as the proportion of patients having colorectal cancer having a parameter representing the concentration of a marker, e.g. TIMP-1 in excreta samples higher than a predefined level.

The method according to the invention can equally well be used for monitoring the response to treatment and the progress of the cancer as rising TIMP-1 values may suggest that a patient could have a negative development of the cancer. In such cases
5 the discriminating value is set specifically for each individual.

Studies have shown that an increase in detection and monitoring values can be obtained when combining the concentration of
10 total TIMP-1 and the concentration of free TIMP-1, thereby providing a highly selective and sensitive method for detecting, screening and monitoring an individual for cancer.

Such cancers could e.g. be breast, prostate, colorectal,
15 cervical, ovarian, lung, pancreatic, renal, vulvar, hepatocellular carcinomas, minimal residual disease and recurrent cancer

In order to improve the sensitivity of the method according to
20 the invention, an additional marker can optionally be employed. Said marker is preferably Carcino Embryonic Antigen (CEA), which is a protein, which is normally found only during foetal development, but may reappear in adults who develop certain types of cancer, primarily cancer of the gastrointestinal
25 system. However, other markers for colorectal cancer known for the person skilled in the art, such as soluble u-PAR, could equally well be employed.

Preferably logistic regression analysis can be used when
30 combining either the concentration of total TIMP-1 and the concentration of free TIMP-1 combination or the combination of concentrations of the cancer marker (CEA) and TIMP-1.

Data has shown that by adding CEA as the additional marker, an
35 improvement in the sensitivity of total TIMP-1 in plasma can be obtained and maintained with a very high specificity. Thus, the

combination of concentrations of CEA and TIMP-1 could be useful in a method for identifying patients with a high risk of having cancer.

- 5 The method according to the invention may be used both for an individual and for an entire population, but more appropriately to a population already identified as having an increased risk of developing cancer, e.g. individuals with a genetic disposition, individuals who have been exposed to carcinogenic substances, or individuals with cancer-predisposing non-malignant diseases.

- 10 As an example this could e.g. be individuals with a prior polyp, individuals with Crohn's disease or ulcerative colitis, individuals with one or more family members with colorectal cancer, or individuals with a prior resection of an early colorectal cancer.

- 20 When an individual has been identified as having high TIMP-1 levels in his or her excreta, the individual should be referred for further examination. If a cancer is found, the patient could be offered surgery, radiation or adjuvant anti-neoplastic therapy aiming at curing the patient of cancer.

- 25 The invention further relates to a dipstick for measurement of the concentration of TIMP-1 either alone or in combination with CEA, in excreta.

- 30 Such a dipstick is extremely easy and quick to use and each individual can perform the test without the need of a professional staff or the use of specialist equipment.

- Individuals in certain high-risk groups, such as individuals having e.g. colitis ulcerosa or individuals with an earlier detected cancer who wants to monitor a possible recurrence of

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the cancer, then have the possibility of screening/monitoring themselves at any desired frequency.

Furthermore, the dipstick can be a valuable tool since each individual has the opportunity to monitor the response to treatment and the spread of cancer without the need for specialist equipment.

The dipstick is preferably a two-sided direct label assay in a lateral flow device format.

In a first embodiment the dipstick can be made of a capillary material, and comprises a first colour indication zone comprising antibodies specific for TIMP-1 and at least one reagent which can give an optically visible colour change in the zone dependent on the concentration of TIMP-1 in the tested excreta.

The dipstick further comprises a second colour indication zone, able to react with at least one substance normally present in the excreta, and thereby providing an optically visible colour change in the zone for controlling that the stick is used properly.

In another embodiment of the dipstick according to the invention, the dipstick further comprises a third colour indication zone, comprising antibodies specific for CEA and at least one reagent which can give an optically visible colour change in the zone dependent on the concentration of CEA in the excreta, ensuring a more reliable screening method.

Preferably there is, in combinations with the dipstick, provided a colour reference that indicates the discrimination value for the specific excreta.

When the user examines the likelihood of having a cancer, the dipstick is first brought into contact with the excreta being tested, e.g. the user can place the dipstick in the mouth thereby bringing it into contact with the patient's saliva.

5

After a certain reaction period determined by the specific reagents on the dipstick, the user can compare the stick with the colour reference scale, dividing the patient into a group with either a high or a low likelihood of having cancer.

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The dipstick can within the scope of the invention be based on any known conventional detection principle based on for instance RIA or enzymatic assays.

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A person skilled in the art would recognise that the dipstick is only one possibility of detecting TIMP-1 in excreta, such as saliva. Other options within the scope of the invention are e.g. an Activity Assay (such as zymography), immunologic assays or a Colour Reaction kit.

20

The invention will be explained in greater detail below, by way of examples only with reference to the drawing, in which

25

Fig. 1 shows the recovery of TIMP-1 signal, measured following addition of increasing concentrations of purified TIMP-1 to a panel of 0.5% and 1% saliva pools in sample dilution buffer,

Fig. 2 shows total TIMP-1 concentration from 39 dilutions of a saliva pool from collection (I) assayed via ELISA, and

30

Fig. 3 shows the TIMP-1 concentration in each saliva sample from collection (I) measurement via ELISA.

35

EXAMPLES:

It should initially be mentioned that even though the concentration of TIMP-1 in both plasma and saliva in the following examples are presented as ng TIMP-1 per volume plasma or saliva, it could equally well be presented as e.g. unit or gram TIMP-1 per unit or gram of a known reference molecule.

Donors/Patients

10 In collection (I), saliva was obtained from four apparently healthy volunteer blood donors.

In collection (II) saliva was obtained from four apparently healthy volunteer blood donors. A simultaneous plasma sample was obtained from each individual at the time of saliva collection.

20 In collection (III) saliva was obtained from three patients with known colon or rectal cancer. A simultaneous plasma sample was obtained from each individual at the time of saliva collection.

Informed consent was obtained from all donors/patients, and permission was obtained from the local Ethical Committees.

25

Saliva collection

In collection (I) saliva was collected from healthy blood donors who were instructed to rinse the mouth with tap water just before the collection. The collection duration was 5 minutes and the donor delivered in this period approximately five saliva samples in a 50 ml Nunc test tube. The saliva is centrifuged 5 minutes at 4000 rpm and the supernatant is then centrifuged at 15000 rpm for 5 minutes.

35 In collection (II) and (III), saliva was collected by introducing a cotton tampon into the mouth for 3 minutes. The

tampon was then centrifuged 5 minutes at 3000 rpm to allow the saliva to be extracted. Subsequently the saliva was centrifuged for 15.000 rpm for 5 minutes.

- 5 The resulting supernatant is in all cases frozen at -20°C, until it is used.

Blood collection and plasma separation

- Peripheral blood was drawn with minimal stasis (if necessary a maximum of 2 min stasis with a tourniquet at maximum +2 kPa was acceptable) into pre-chilled citrate, EDTA, or heparin collection tubes (Becton-Dickinson, Mountain View, CA), mixed 5 times by inversion, and immediately chilled on ice. As soon as possible (no later than 1.5 h after collection) the plasma and blood cells were separated by centrifugation at 4°C at 1,200 x g for 30 min, and stored frozen at -80°C prior to assay. Plasma pools were made with freshly collected samples from at least ten donors, aliquoted and stored frozen at -80°C. For analysis, the samples were quickly thawed in a 37°C water bath and placed on ice until needed.

Example 1: Measurement of total TIMP-1 concentration in saliva

- Preparation of an ELISA to quantitate total TIMP-1 concentrations in human plasma.

- A sensitive and specific sandwich ELISA was prepared, using TIMP-1 antibodies developed at the Strangeways Laboratories (Hembry et al, 1985). A sheep polyclonal anti-TIMP-1 antiserum (Hembry et al, 1985; Murphy et al, 1991) was used for antigen capture, and a murine monoclonal anti-TIMP-1 IgG1 (MAC-15) (Cooksley et al, 1990) for antigen detection.

- A rabbit anti-mouse immunoglobulin/alkaline phosphatase conjugate (Catalog number D0314, Dako, Glostrup, Denmark) was the secondary detection reagent. The latter conjugate was supplied

preabsorbed against human IgG, thus eliminating cross-reactivity with IgG in the samples.

5 As the monoclonal detection antibody MAC-15 recognises both free TIMP-1 and TIMP-1 in complex with MMP's (Cooksley et al, 1990), the total TIMP-1 content captured by the sheep polyclonal anti-TIMP-1 antiserum was quantitated by the ELISA.

96-well microtiter plates (Maxisorp, Nunc, Roskilde, Denmark)
10 were coated for 1 h at 37°C with 100 µL/well of polyclonal sheep anti-TIMP-1 (4 mg/L) in 0.1 mol/L carbonate buffer, pH 9.5. The wells were then rinsed twice with 200 µL/well of SuperBlockJ solution (Pierce Chemicals, Rockford, IL) diluted 1:1 with phosphate-buffered saline (PBS). The microtiter plates
15 were stored for up to 14 days at -20°C. On the day of analysis, the plates were thawed at room temperature and washed 5 times in PBS containing 1 g/L Tween.

A series of purified, recombinant human TIMP-1 standards were
20 used to calibrate each plate. Standards were prepared by serially diluting a stock solution of purified TIMP-1. Standard concentrations were 5, 3, 2, 1, 0.5, 0.25, 0.1, ~0 ng/mL. Included on each plate was a blank containing only sample dilution buffer, and 2 controls made from a 1:100 dilution of a
25 citrate plasma pool. One control was added as the first sample on the plate and the second control was added as the last.

All samples were diluted 1:100 or 1:51 in sample buffer consisting of 50 mol/L phosphate, pH 7.2, 0.1 mol/L NaCl, 10
30 g/L bovine serum albumin (Fraction V, Boehringer-Mannheim, Penzberg, Germany), and 1 g/L Tween 20.

A total of 100 µL/well of each standard, blank, control, and patient sample was incubated on the plate for 1 h at 30°C. All

standards, blanks, controls, and samples were run in triplicate on each plate for every assay.

After primary incubation, the wells were washed 5 times, then treated for 1 h at 30°C with 100 µL/well of purified MAC-15 monoclonal antibody (0.5 mg/L) in sample dilution buffer. After another 5 washes the wells were incubated for 1 h at 30°C with 100 µL/well of rabbit anti-mouse immunoglobulins(Ig)/alkaline phosphatase conjugate diluted 1:2000 in sample dilution buffer.

Following 5 washes with washing solution and 3 washes with distilled water, 100 µL of freshly made p-nitrophenyl phosphate (Sigma, St. Louis, MO) substrate solution (1.7 g/L in 0.1 mol/L Tris.HCl, pH 9.5, 0.1 mol/L NaCl, 5 mmol/L MgCl₂) were added to each well.

The plate was placed in a Ceres 900J plate reader (Bio-Tek Instruments, Winooski, VT) at 23°C with the yellow colour development automatically monitored. Readings were taken at 405 nm against an air blank every 10 min. for one hour. KinetiCalc II software was used to analyse the data by calculating the rate of colour formation for each well (linear regression analysis), generating a 4-parameter fitted standard curve, and calculating the TIMP-1 concentration of each sample.

Recovery experiments

The recovery of the TIMP-1 signal was measured following addition of increasing concentrations of purified TIMP-1 to a panel of 0.5% and 1% saliva pools in sample dilution buffer, followed by subsequent measurement of the signal.

The purified TIMP-1 was added to dilution series of the saliva pools to give concentrations in the range of 0 to 5 ng/mL.

Discussion

The assay described above enables accurate determination of total TIMP-1 in human saliva samples. The use of a rapid blocking agent and a dilution buffer with high buffering capacity also contributed to reproducible assays. Incorporating all these elements in the final assay fulfilled the requirements of sensitivity, specificity, stability, and good recovery of an internal standard.

The quantitative studies in saliva from healthy donors showed that saliva samples are suitable for TIMP-1 determination.

These studies showed that for collection (I) TIMP-1 levels in saliva (mean 72-79 ng/mL) correspond to TIMP-1 levels in plasma (mean 65-70 ng/mL) (see International Patent Application No. WO00/62070). Similar results were obtained from the healthy donors in collection (II).

Example 2: Detection value of total TIMP-1 in patients with colorectal cancer.

Total TIMP-1 levels in both plasma and saliva from three colorectal cancer patients (collection (III)) were measured with the TIMP-1 assay described in Example 1. The TIMP-1 values were analyzed and compared using standard biostatistical parameters.

Patients

Three patients undergoing elective surgery for pathologically confirmed colorectal cancer were included in the study. Blood and saliva samples were obtained preoperatively with informed consent from all patients in accordance with the Helsinki declaration, and permission was granted by the local Ethical Committee of Hvidovre Hospital, Denmark. All patients had pathologically verified adenocarcinoma of the colon or rectum.

Measurement of Total TIMP-1 concentration in collection (III)

In collection (III) both the saliva samples and the plasma samples from colorectal patients were assayed for total TIMP-1 concentration via ELISA. The results are presented in table 2:

Colorectal patient	Total TIMP-1 concentration in saliva (ng/mL)	Total TIMP-1 concentration in EDTA plasma (ng/mL)
5	328	395,7
6	409,9	127
7	358,9	172,3

Table 2: Total TIMP-1 concentration in saliva and plasma in ng/mL in three colorectal patients.

Discussion

15 This data proved that similar saliva/plasma TIMP-1 concentrations were found not only for healthy individuals but also in patients suffering from colorectal cancer.

20 Furthermore, all three colorectal cancer patients had elevated (above the 95 percentile of healthy blood donor plasma levels) saliva and plasma TIMP-1 levels. There is therefore a highly statistical difference in the total saliva TIMP-1 values between the colorectal cancer patients and the healthy blood donors.

25 It has previously been found, that a highly statistical difference in the total plasma TIMP-1 values between the colon and rectal cancer patients each compared with the healthy blood donors exists. Furthermore, it has been shown in the
30 aforementioned examples that corresponding concentrations of total TIMP-1 is present in saliva and plasma. The person skilled in the art would based on these finding therefore expect that the same statistical difference between the colon and rectal cancer patients would be observed in total TIMP-1
35 from saliva.

10

30 The optimal discriminating value for TIMP-1 in saliva to
discriminate between malignant and normal tissues can be
determined on the basis of such ROC curves as the point on the
curve, which shows the optimal combination for specificity and
sensitivity and where diagnostic accuracy is maximized. The
35 discriminating value for total TIMP-1 in plasma based on 870
healthy blood donors has been determined (Brünner, unpublished

data), and based on the above findings a similar result is expected for total TIMP-1 in saliva.

Example 3: Measurement of the total concentration of TIMP-1:MMP-9 complexes and free TIMP-1 in saliva

As shown in Example 1 the total TIMP-1 concentration in saliva corresponds very accurately to the total TIMP-1 concentration in plasma. As it has been found that there exists a correlation between plasma and saliva, it would be obvious to the person skilled in the art to also examine the total concentration of TIMP-1:MMP-9 complexes and free TIMP-1 concentration in saliva. Appropriate methods and materials are described in similar examples in WO00/62070 said reference being incorporated herein by reference. In view of the findings in Example 1, it would be expected that similar results would be obtained for the total concentration of TIMP-1:MMP-9 complexes and free TIMP-1, respectively.

Example 4: Detection value of total TIMP-1 in combination with CEA in patients with colorectal cancer.

Examples in WO00/62070 have demonstrated that by adding an additional marker, an improvement in the diagnostic sensitivity of total TIMP-1 in plasma can be obtained, while maintaining a high specificity of 98%. Thus, based on the assumption that corresponding data can be obtained with total TIMP-1 in plasma and saliva, the combination of CEA and TIMP-1 in saliva could be useful as a screening procedure to identify patients with a high risk of having colorectal cancer.

Example 5: Lack of detection value of total TIMP-1 in saliva in patients with primary (stage I and II) breast cancer.

Using the total TIMP-1 assay described in Example 1, total TIMP-1 levels in pre-operative plasma samples from 322 patients

with primary breast cancer were compared with 108 plasma samples from healthy blood donors. Data shown in WO00/62070, which is incorporated herein by reference, showed no statistical significant difference in total TIMP-1 plasma levels between the two groups (Mann-Whitney, $p=0.87$). Thus, this data supports the specificity of TIMP-1 measurements in the diagnosis of patients with colorectal cancer. Similar results would be expected if the total TIMP-1 level were measured in saliva samples from similar donors.

10

Example 6: Detection value of saliva total TIMP-1 in patients with metastatic (stage IV) breast cancer.

Based on information from WO00/62070 where total TIMP-1 levels were measured in 19 breast cancer patients with stage IV disease all EDTA plasma samples using the assay described in Example 1. These levels were compared with total TIMP-1 plasma levels in 87 healthy female donors. A Wald-Wolfowitz test indicated a highly significant difference ($p<0.0001$) between patient total TIMP-1 levels and those of healthy donors. As it has been established that corresponding levels of total TIMP-1 are present in plasma and saliva these data show that saliva TIMP-1 measurements can be used to monitor breast cancer patients for recurrence of disease.

25

Example 7: Detection value of free TIMP-1 in saliva in patients with colorectal cancer.

In WO00/62070 it was shown that free TIMP-1 in plasma alone is not likely to be useful as a screening marker to identify patients with a high risk of having colorectal cancer. As previously mentioned based on the results shown in the present application similar results are expected for free TIMP-1 in saliva.

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Example 8: Screening value of a dipstick for detection of TIMP-1 in saliva.

Procedure

5 A colloidal gold/antibody conjugate was produced for practice of the methods of the present invention. Siliconized glassware (Sigma silicote) was utilized throughout the procedure wherein 200 ml of 0.01% gold chloride ($\text{HAuCl}_4 \cdot 3\text{H}_2\text{O}$) (Fisher Scientific, G-54-1) was brought to a boil and 2 ml of 1% sodium
10 citrate solution was added and the boiling is continued for 5 minutes until the color of the solution changes from pale yellow to purple to red. A solution of potassium carbonate (0.02M) was added to the suspension in order to adjust the pH to 7.6, followed by addition of the rabbit polyclonal anti-MMP-
15 9 antibody (1 mg/ml) (Novus Biologicals Product) such that approximately 10 μg IgG was added per ml of gold suspension (0.01% gold).

After one minute of incubation at room temperature, 0.1 ml of a
20 solution of 30% bovine serum albumin in water was added to 10 ml of the gold suspension. Aggregated material was removed by centrifugation at 3000 rpm in the SS34 rotor of a Sorval RC5C centrifuge for 10 minutes. The supernatant was subjected to an additional centrifugation step at 6000 rpm for one hour. The
25 colloidal gold conjugate in the pellet was resuspended in 2% bovine serum albumin in PBS (0.05 M potassium phosphate buffer, pH 7.4, in 0.9% NaCl), the preferred conditions for liquid storage being at 4°C. Casein solution was added to give a concentration of 1% casein immediately before application to
30 the membrane. The conjugate was then stored for prolonged periods in the dry state, with no loss of activity after 6 months storage at 37° C. in the dry state.

According to this example, sandwich-type immunoassay devices
35 for the detection of TIMP-1 were constructed and used. Microporous nitrocellulose material with a thickness of

approximately 0.1 mm and an average pore size of 5 μ m was laminated with mylar and adhesive (Monokote, Top Flite Models, Inc., Chicago, Ill.). Strips measuring 1 cm by 3.5 cm were cut. Mouse monoclonal antibody MAC 19 (see WO/62070), 0.2 μ l, was applied to the strips at the capture zone where it was immobilized and air dried. Non-specific binding sites on the chromatographic strip materials were then blocked by incubation for 10 minutes at room temperature with a 0.1% solution of LB gelatin in water (Inotech, Wohlen, Switzerland) and the strips were allowed to dry under a stream of air. One μ l of gold particle labeled rabbit polyclonal anti-MMP-9 antibody in an anti-aggregation buffer produced as described was then applied to a first zone of each strip and dried.

15 Detection of TIMP-1 concentration in saliva samples on a dipstick

Saliva samples from collection (II) and (III), healthy blood donors and colorectal cancer patients, respectively, were first applied to the second zones between the first and third zones.

20 The first end of the strips were then dipped into a chromatographic transport solvent comprising TBS and 1% TRITON X-100 (polyethylene glycol tert-octylphenyl ether). The liquid front was allowed to progress to the second ends of the devices over a period of approximately 2.5 minutes transporting the sample material and the gold-labeled goat anti-human IgG to the third zone.

30 Samples from the colorectal cancer patients (collection (III)) and the immobilization of the labeled first reagent resulted in the presence of a red spot at the third zone. Strips tested with samples from the healthy blood donors, collection (II) did not produce a signal at the third zone.

Discussion

These test confirm that TIMP-1 can be measured in saliva by means of a dipstick; and that such a dipstick can be used in a screening procedure to aid in identifying patients with a high risk of having colorectal cancer. Furthermore, the dipstick can be used to monitor the response to treatment and the spread of cancer without the need for specialist equipment.

CONCLUSION

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The person skilled in the art would understand from the present invention that the similarities between total TIMP-1 levels in plasma and saliva is not limited to the Examples mentioned in the current application. In fact it is now surprisingly expected that for any TIMP-1 value measured in a plasma sample a corresponding value could be established for TIMP-1 in saliva.

20 This is not just true for the examples mentioned in the present application or in WO00/62070 but also for all other known results obtained for plasma.

25 The data presented in the present application supports the unique value of testing saliva when screening, monitoring and detecting early stage colorectal cancer or metastatic breast cancer using TIMP-1 as a cancer marker. The method for identification of patients having primary colorectal cancer is highly specific, and patients with non-malignant conditions, such as inflammatory bowel diseases, are not detected. 30 Furthermore, saliva may also be suitable when monitoring the response to treatment and the progress of cancer.

The description has primarily described excreta from humans; however it is within the scope of the invention that the excreta is from an animal, e.g. a dog, cat or cow. 35

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21.

population with a predetermined specificity or a predetermined sensitivity.

5 8. A method according to any of the preceding claims, wherein the method further comprises determining at least one second parameter representing the concentration of a marker for cancer different from any form of TIMP-1, in an excreta from an individual.

10 9. A method according to claim 8, wherein the first and second parameter are combined to result in a combined parameter wherein the presence of a concentration of the combined parameter above a predetermined discrimination value is an indication that the individual has a high
15 likelihood of having a cancer or there being a progression in a cancer.

20 10. A method according to claim 9, wherein the discrimination value is determined by determining the combined parameter in the at least one excreta in both a healthy control population and a population with known colorectal cancer, thereby determining the discriminating value which identifies the cancer population with a predetermined specificity or a
25 predetermined sensitivity.

30 11. A method according to claim 9 or 10, wherein the combination of the first and second parameter is performed by logistic regression analysis.

12. A method according to claim 8-11 wherein the at least one second parameter is the concentration of Carcino Embryonic Antigen (CEA).

13. A method according to any of the preceding claims, wherein the determination of the concentration is performed by means of an immunoassay or an active assay.
14. A method according to any of the preceding claims, wherein the immunoassay is an ELISA.
15. A method according to any of the preceding claims, wherein the active assay is zymography.
16. Use of a method according to claim 1-15, for detection of early stage cancer.
17. Use according to claim 16, for detection of early stage colorectal cancer.
18. Use according to claim 16, for detection of metastatic breast cancer.
19. Use of a method according to claim 1-15, for monitoring the response to cancer treatment.
20. Use of a method according to claim 1-15, for monitoring the recurrence of a cancer.
21. A dipstick for performing the method according to claim 1-15, wherein said dipstick comprises a first colour indication zone, comprising antibodies specific for TIMP-1.
22. A dipstick according to claim 21, wherein the first zone further comprises at least one reagent which can give an optically visible colour change in the zone dependent on the concentration of TIMP-1 in at least one excreta.

23. A dipstick according to claim 21 or 22, wherein the dipstick further comprises a second colour indication zone, able to react with at least one substance normally present in the excreta, and thereby providing an optically visible colour change in the zone for controlling that the stick is used properly.
24. A dipstick according to claim 21-23, wherein said dipstick further comprises a third colour indication zone, comprising antibodies specific for CEA.
25. A dipstick according to claim 21-24, wherein the first zone further comprises at least one reagent which can give an optically visible colour change in the zone dependent on the concentration of CEA in at least one excreta.

1/3

Recovery experiment for TIMP-1 concentration in saliva
for collection (I).

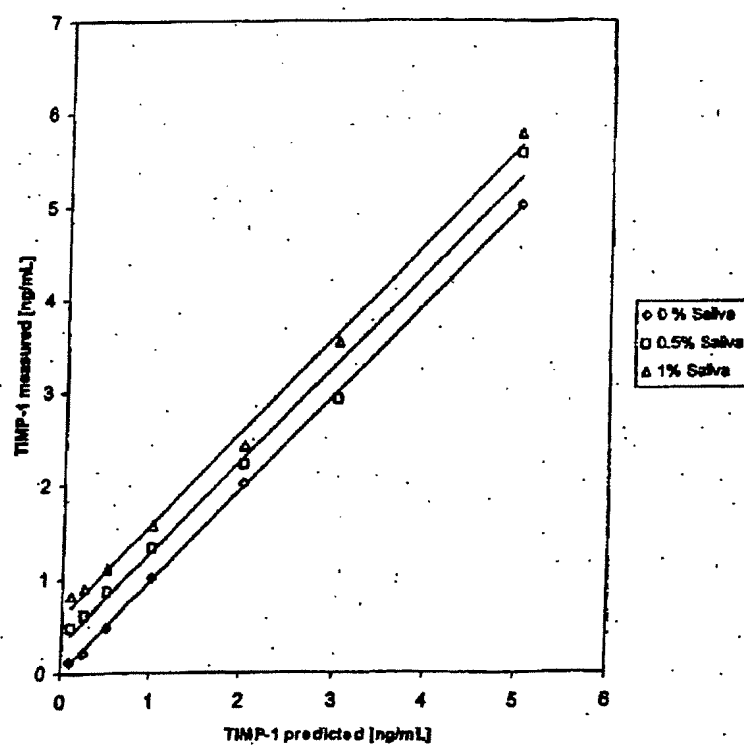


Figure 1.

2/3

Measurement of TIMP-1 concentrations in a saliva pool
from collection (I).

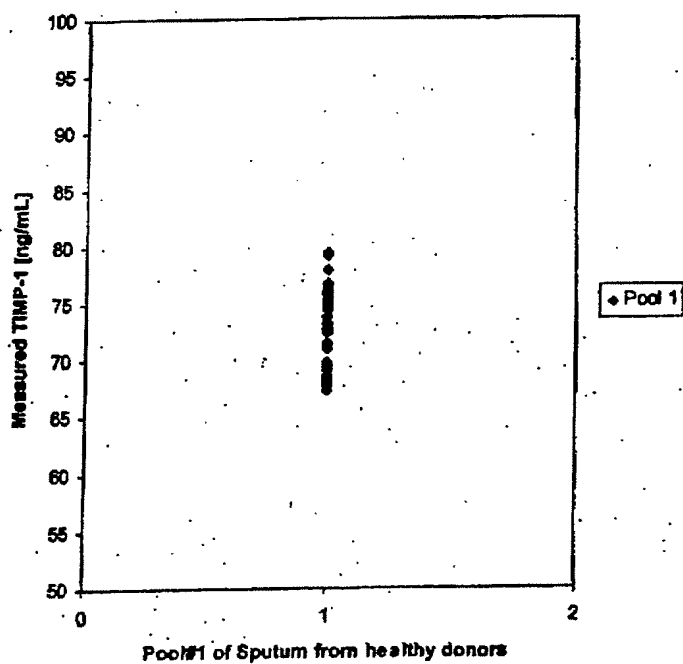


Figure 2.

3/3

Measurement of TIMP-1 concentration in different donors from collection (I).

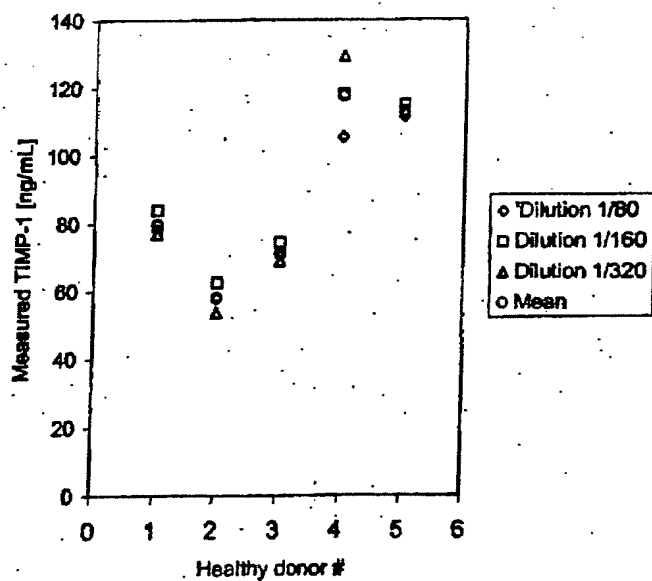


Figure 3.

DECLARATION AND POWER OF ATTORNEY FOR PATENT APPLICATION

As the below named inventor, I hereby declare that:

My residence, post office address and citizenship is as stated below next to my name;

I believe that I am the original, first and sole inventor (if only one name is listed below) or an original, first and joint inventor (if plural names are listed below) of the subject matter which is claimed and for which a patent is sought on the invention entitled:

A METHOD FOR DETECTING, SCREENING AND/OR MONITORING A CANCER IN AN INDIVIDUAL

the specification of which:

☐ is attached hereto.
☒ was filed on: September 26, 2003
as United States Application No.: _____
or PCT International Application No.: PCT/DK2003/000634
and was amended on: March 28, 2005 (if applicable).

I hereby state that I have reviewed and understand the contents of the above-identified specification, including the claims, as amended by any amendment specifically referred to above.

I acknowledge the duty to disclose information which is material to patentability as defined in 37 C.F.R. § 1.56.

Prior Foreign Application(s)

I hereby claim foreign priority benefits under 35 U.S.C. §§ 119(a)-(d) or (f), or § 365(b) of any foreign application(s) for patent, inventor's or plant breeder's rights certificate(s), or § 365(a) of any PCT international application which designated at least one country other than the United States of America, listed below and have also identified below by checking the box, any foreign application(s) for patent, inventor's or plant breeder's rights certificate(s), or any PCT international application having a filing date before that of the application on which priority is claimed:

Application Number	Country	Date of Filing (day, month, year)	Date of Issue (day, month, year)	Priority Claimed	
PA 200201430	Denmark	26-09-2002		Yes <input checked="" type="checkbox"/>	No <input type="checkbox"/>
DK 199900476	Denmark	09-04-1999		Yes <input checked="" type="checkbox"/>	No <input type="checkbox"/>

Prior Provisional Application(s)

I hereby claim the benefit under 35 U.S.C. § 119(e) of any United States provisional application(s) listed below:

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